# Microgravity and the implications for wound healing

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Farahani RM, DiPietro LA. Microgravity and the implications for wound healing. Int Wound J 2008;5:552-561.

### **ABSTRACT**

Wound healing is a sophisticated response ubiquitous to various traumatic stimuli leading to an anatomical/ functional disruption. The aim of present article was to review the current evidence regarding the effects of microgravity on wound healing dynamics. Modulation of haemostatic phase because of alteration of platelet quantity and function seems probable. Furthermore, production of growth factors that are released from activated platelets and infiltration/function of inflammatory cells seem to be impaired by microgravity. Proliferation of damaged structures is dependent on orchestrated function of various growth factors, for example transforming growth factors, platelet-derived growth factor and epidermal growth factor, all of which are affected by microgravitational status. Moreover, gravity-induced alterations of gap junction, neural inputs, and cell populations have been reported. It may be concluded that different cellular and extracellular element involved in the healing response are modified through effect of microgravity which may lead to impairment in healing dynamics.

**Key words:** Cytokine ullet Growth factor ullet Microgravity ullet Wound healing

# **Key Points**

- regardless of the specific aetiology and external manifestation, the similar array of events that follows the injury is directed at restoration of the original status of wounded tissue
- considering the probability that traumatic injuries will occur during space travel and the importance of efficient repair to the astronaut, it will be crucial to understand the impact of microgravity on various elements and phases of wound healing
- in the present article, we review the current information about the modulation of healing by gravity

### INTRODUCTION

Wounds involve the gross macroscopic or subliminal microscopic damage to the anatomical and functional integrity of live tissues.(1,2) The diverse clinical manifestations of injury range from conspicuous cutaneous injuries (3) to scenarios involving subtle metabolic microangiopathies (4). Regardless of the specific aetiology and external manifestation, the similar array of events that follows the injury is directed at restoration of the original status of wounded tissue. The repair process progresses through several overlapping phases, which include inflammation, proliferation and remodelling (2,3). Alteration of any of the interrelated elements involved in healing can affect the entire process and the final outcome (5–8).

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The modern era has witnessed a substantial rise in quantity and quality of manned space flights and has provided promise for the eventual long-term inhabitation of space, either on stations or other planets. Within space, the dynamic equilibrium of human body is altered by exposure to a variety of altered circumstances (9,10). These include changes in gravitational status, neuroimmunoendocrine modulations, modified environmental stimuli (such as radiation), etc. (10). Considering the probability that traumatic injuries will occur during space travel and the importance of efficient repair to the astronaut, it will be crucial to understand the impact of microgravity on various elements and phases of wound healing. In the present article, we review the current information about the modulation of healing by gravity.

### Haemostasis

Immediately after wounding, two interrelated pathways are activated in order to stop bleeding. A fibrin clot forms, filling the anatomical void, while platelets contact the exposed collagen and undergo aggregation and activation. The resulting fibrin/platelet plug contributes to the initial stability of wound and also serves as a provisional matrix that will be replaced as healing proceeds (11,12). Beyond their haemostatic function, platelets release several growth factors such as transforming growth factor-beta (TGF- $\beta$ ), platelet-derived growth factor (PDGF), insulin-like growth factors and epidermal growth factor (EGF), all of which may play a role in the initiation of healing response (13,14).

Studies that examine the effects of microgravity on haemostatic pathways are contradictory, as reports of both increased thrombotic disease (15) and increased haemorrhagic status (16) can be found in the extant literature. Thrombocytopenia, an increase in activated partial thromboplastin time and defective cell-cell adhesion have been described to occur in the microgravitational condition, increasing the haemorrhagic risk. The sustained elevation of blood pressure that has been shown to result because of microgravity exposure may also increase the threat of haemorrhage (17-21). In contrast, in circumstances of altered gravity, thrombotic incidents may increase because of alterations in blood volume, blood viscosity, elevation of superoxide anions, increases in catecholamine release and metabolic disorders (22-25). Coagulative imbalances have also been reported (26). Fibrin structure, including the thickness of the fibres, the number of branch points, porosity and permeability, have also been suggested to be important determinants of healing outcomes (12). It has been reported that microgravityformed fibrin gels are more uniform than those formed at normal gravity, although the fibre diameter and matrix porosity are not affected (27) In contrast, later studies showed diminished matrix porosity in microgravitational status (28). The relative inconsistency of data may partially reflect the difference in experimental methods and environments. One possibility is that the diminished platelet content of the platelet/fibrin plug owing to microgravity may reduce contractile force and promote the fibrinolysis rate of fibrin clot (29–31). An earlier loss of superficial eschar because of hastened fibrinolysis would be expected to lead to increased wound contraction (32). Independent of whether haemostasis is altered, bleeding from the wound site under situations of microgravity would permit the formation of large fluid dome because of high surface tension of blood (33).

While control of this bleeding pattern may not be difficult (33), special training for providing medical care to wounds may be needed because of altered fine motor skill (34).

### Inflammation

Degranulation of activated platelets is an early event in wound healing. Platelets release a multitude of potential mediators at the wound site, including TGF-β, PDGF and EGF, and many of these mediators may modulate the subsequent healing process. PDGF can initiate the chemotaxis of neutrophils, smooth muscle cells and fibroblasts. This growth factor is mitogenic for fibroblasts and arterial smooth muscle cells, can facilitate the release of other growth factors and is also reported to enhance the effects of some growth factors like TGF-β (35–37). Similarly, TGF- $\beta$  isoforms (TGF- $\beta$ 1, TGF- $\beta$ 2 and TGF- $\beta$ 3) can attract neutrophils to sites of injury (38). The most important role of TGF-β is probably the regulation of the deposition of extracellular matrix components, which occurs through its influence on the proliferation of fibroblasts and their synthetic activity (39). Moreover, TGF-β and PDGF enhance the effect of each other through synergistic interactions (36,37,40). A similar synergistic interaction has been reported for EGF and PDGF (41). While platelets certainly provide an initial pool of EGF, TGF-β and PDGF, other cell types within the wound continue to produce these mediators as healing progresses (42–50).

Space flight probably has the potential to greatly alter the production of EGF, PDGF and TGF- $\beta$  in wounds. The microgravitational status has been shown to influence the expression the EGF receptor, and EGF-induced signal transduction is impaired in microgravity (51-53). The production of TGF-β has been found to be downregulated in simulated microgravitational status, and microgravity induces the downregulation of the PDGF receptor by 62% (54). Furthermore, the response of wounds to PDGF during space flight was attenuated compared with control wounds on the ground (55). Together, these data suggest that the production of growth factors that are released from activated platelets seems to be impaired by microgravity.

The inflammatory phase involves the regional activation of immune system and the infiltration of wound site by inflammatory cells. Many of the infiltrating immune cells secrete growth factors that stimulate the proliferation of cellular components of the tissues. Neutrophils

# **Key Points**

- studies that examine the effects of microgravity on haemostatic pathways are contradictory, as reports of both increased thrombotic disease and increased haemorrhagic status can be found in the extant literature
- the relative inconsistency of data may partially reflect the difference in experimental methods and environments
- independent of whether haemostasis is altered, bleeding from the wound site under situations of microgravity would permit the formation of large fluid dome because of high surface tension of blood
- while control of this bleeding pattern may not be difficult, special training for providing medical care to wounds may be needed because of altered fine motor skill
- space flight probably has the potential to greatly alter the production of EGF, PDGF and TGF-b in wounds
- these data suggest that the production of growth factors that are released from activated platelets seems to be impaired by microgravity

# **Key Points**

- beyond cell number, functional attributes of leucocytes that are critical to cell migration also appear to be altered by space flight
- several studies suggest that mast cells influence inflammation and repair at sites of injury and a link between increased mast cell content and the formation of hypertrophic scars has been suggested
- comprehensive research aiming at the elucidation of the function and number of these cells in microgravity-exposed wounds seems necessary

**Table 1** The effect of microgravity on various elements involved in wound healing

Target	Increase	Decrease	Unaffected
Platelets		+ (16)	
Superoxide anions	+ (22-25)		
Fibrin porosity		+ (28)	+ (27)
Fibrinolysis	+ (29-31)		
EGF function		+ (51-53)	
TGF-β		+ (54,85,86)	+ (87)
PDGF function		+ (54,55)	
Monocytes		+ (59)	
Neutrophils		+ (60,61)	+ (62)
T cell		+ (72–75)	
IL-2		+ (72,74,80)	
IL-1β		+ (79)	
IL-6	+ (80-81)		
TGF- $\alpha$		+ (100-103)	
Gap junctions		+ (113)	+ (114)
Stem cell activity	+ (129)	+ (127–129)	

EGF, epidermal growth factor; TGF, transforming growth factor; IL. interleukin.

are the first of the circulating inflammatory cells to arrive at the site of injury (56,57). Thereafter, monocytes gradually arrive, eventually becoming the dominant inflammatory cell population in wounded region. It has been suggested that the abundance of monocytes, precursors of macrophages, appears to be a rate-limiting parameter in tissue repair (58). In this context, space-induced deficiencies in monocytes may be critical. Indeed, Taylor et al. (59) reported a decrease in peripheral blood monocytes after space flight. Likewise, Ichiki et al. (60) reported microgravity-induced neutrophilia. This finding may partially reflect the effect of flight-induced psychological stress on immune function (61). In contrast, Meehan et al. (62) reported a post-flight increase in circulating monocytes and no significant changes in plasma cortisol levels. Allebban et al. (63) reported a significant reduction in the absolute number of lymphocytes and monocytes and a slight increase in the absolute number of eosinophils and neutrophils after space flight. Thus a microgravity-induced reduction in monocytes seems a consistent finding, while changes in the number of neutrophils are less consistently reported.

Beyond cell number, functional attributes of leucocytes that are critical to cell migration also appear to be altered by space flight. While the studies in this area are somewhat conflicting, overall they provide strong evidence that microgravity does indeed influence leucocyte function. Most of the data support the concept that neutrophil adhesiveness is increased by microgravity. The levels of adhesion molecules on neutrophils are increased during space flight (64), and a 10-fold increase in chemotactic response of neutrophils exposed to microgravity has been shown. The findings of Boxer et al. (65), who showed impaired locomotion of monocytes in modelled microgravity, suggest that the effect is cell specific. Mechanistically, flight stress, and the resulting catecholamine release, may be important to these observed changes in leucocyte function. However, direct exposure of neutrophils to epinephrine had no direct effect on neutrophil adhesion (66); although exposure of endothelial cells to epinephrine decreased neutrophil adherence by 40%, the oxidative functions and microgravity are less well investigated. However, an antiorthostatic suspension (modelled space flight) did not alter the oxidative burst in neutrophils (67).

Another immune cell that might be influenced by space flight is the mast cell. Mast cells release vasoactive amines, which enhance the permeability of regional blood vessels, promoting the passage of solutes and inflammatory cells to wound site. Several studies suggest that mast cells influence inflammation and repair at sites of injury and a link between increased mast cell content and the formation of hypertrophic scars has been suggested (68). To date, the effect of gravity on mast cell function has not been evaluated. However, stress seems likely to affect mast cell function, as steroid-therapy reduces the number of these cells in hypertrophic scars (69). Psychological stress in rats resulted in dura mast cell activation and rat mast cell protease I secretion that were corticotropin-releasing hormone (CRH) dependent (70). Also, it has been proposed that CRH activates skin mast cells leading to vasodilation and increased vascular permeability (71). Although speculative, flightinduced psychological stress might be predicted to cause a sustained hyperactivation of these cells in the wound milieu, with resultant increased hypertrophic scarring. Nonetheless, comprehensive research aiming at the elucidation of the function and number of these cells in microgravity-exposed wounds seems necessary.

Another immune cell type that appears to be functionally impaired by exposure to microgravity is Tcells (72–75). The expression of both IL-2 and IL-2Ralpha genes is significantly inhibited in simulated microgravity (73).

Moreover, purified human T lymphocytes are shown to exhibit differential inhibition of transcription factor activation in modelled microgravity. Activation of Activator Protein 1 (AP-1) is blocked with clinorotation, whereas dephosphorylation of nuclear factor of activated T cells occurs (75). While the exact role of T cells in regeneration of damaged tissues is not known, space-induced functional impairments may need to be considered in the context of wounds.

Beyond direct effects on cells themselves, microgravitational exposure may influence the production of the cytokines that connect the cellular elements of wound milieu. Peana et al. (76) assessed the effect of microgravity on Prostaglandin E2 (PGE2)-induced oedema and hyperalgesia. Both oedema and hyperalgesia decreased because of anti-inflammatory and anti-hyperalgesic action of simulated microgravity. In contrast, Kumei et al. (77) detected enhanced levels of PGE2 in flight samples compared with ground controls in normal rat osteoblast cultures. The secretion of interleukin (IL)-1β, a factor that exerts panoply of effects in wound milieu (78), is almost completely inhibited in microgravity (79). IL-1 is a potent inducer of IL-6. However, despite inhibition of IL-1, the level of IL-6 increased during space flight (80,81). In one report, the expression of IL-2 and IL-2 receptor was significantly suppressed at simulated zero gravity (72,80), although another report failed to detect any alteration of IL-2 level at zero gravity (73). Overall, the available studies of the influence of gravitational stress on cytokine production suggest that microgravity may produce multiple perturbations in secretory patterns at sites of inflammation, such as the healing wound.

# Proliferation

In skin, the proliferative stage of healing involves regeneration of epithelial barrier, deposition of extracellular matrix and proliferation of connective tissue cells. Various growth factors secreted during inflammatory stage mediate differentiation and proliferation and function of cellular elements. One key factor is TGF- $\beta$ , which regulates the deposition of new extracellular matrix through transcriptional activation of genes encoding extracellular matrix molecules such as collagen and proteoglycans (82). TGF- $\beta$ , can also inhibit tissue protease production and stimulate the secretion of the inhibitors of matrix metalloproteinases

(83,84). The majority of the available data indicates that expression of the various isoforms of TGF-β is reduced by exposure to microgravity (85,86), although one study found no such effect (87). The tissue response to TGF-β is decreased under microgravity, suggesting impairment in signal transduction pathways (88). As mentioned above, the influence of space on cytokines such as TGF-β may be simply an indirect effect that stems from flight-induced psychological stress. In support of this concept, glucocorticoids are known to antagonise the effect of TGF- $\beta$  at the level of transcription (89). Song et al. (90) found that glucocorticoids repress TGF-β activation of the TGF-β responsive sequence containing Smad3/4-binding sites. Biomechanical properties of tissue may also influence the modulation of TGF- $\beta$  function in microgravity, as microgravity generates a low shear strain environment. While shear stress enhances the expression of TGF-β, blocking this mechanical stimulus inhibits its expression (90). Hence, it may be expected that gravity-induced fluid dynamics may lead to downregulation of TGF-β. While extrapolation of these results to regenerating tissues may not be direct, it seems likely that microgravity and the stress of space travel will result in an impairment of both TGF- $\beta$  production and the cellular response to this cytokine. Given that mice deficient in TGF-β or its signalling components exhibit significant deficits in healing (91), space flight-induced alterations in TGF-β or its signalling pathways would probably have extreme effects on healing. It is known that the biological activity of TGF-β depends mainly on the type of activated receptor/signal transduction pathway and to a lesser extent on the specific isoforms involved. Therefore, we suggest the use of knockout models in microgravity studies of wound healing to efficiently target the alteration in TGF-β signalling cascade and associated modulation of regenerative outcome.

The EGF family, which includes EGF, transforming growth factor- $\alpha$  (TGF- $\alpha$ ) and heparinbinding EGF, also seems to be influenced by microgravity (92,93). Several studies suggested that EGFR is important for reepithelialisation, especially during early stages of healing (94,95). EGF and TGF- $\alpha$  appear to be critical to the development of the normal phenotypic features of regenerating epithelium (96,97). However, the deficiency of these growth factors has been reported to be compensated by other growth

# **Key Points**

- beyond direct effects on cells themselves, microgravitational exposure may influence the production of the cytokines that connect the cellular elements of wound milieu
- the available studies of the influence of gravitational stress on cytokine production suggest that microgravity may produce multiple perturbations in secretory patterns at sites of inflammation, such as the healing wound
- the tissue response to TGF-b is decreased under microgravity, suggesting impairment in signal transduction pathways
- we suggest the use of knockoutmodels in microgravity studies of wound healing to efficiently target the alteration in TGF-b signalling cascade and associated modulation of regenerative outcome

# **Key Points**

- the expression of EGF and TGFa is substantially down regulated and the signal transduction pathways are impaired in microgravity
- space flight-induced psychological stress may lead to down regulation of PDGF and its receptor in wound milieu
- following trauma, rapid immediate closure of gap junctions takes places, uncoupling damaged cells from uninjured ones-directional centripetal migration of keratinocytes into the wound bed is necessary for optimal wound healing
- while sympathetic nervous activity was decreased during head-down bed rest, a similar finding was not observed during microgravity and thus headdown bedrest cannot be applied to simulate changes in sympathoadrenal activity during microgravity
- considering the existing controversy, investigation of microgravity-induced alteration of sympathetic activity and its effect on healing dermal wounds seems necessary
- the contribution of epidermal stem cells to repair of wounded epidermis is now evident
- newly emerging data contradict the proposed negative impacts of microgravity and underline its enhancement of proliferative activity of mesenchymal stem cells

factors such as keratinocyte growth factor (KGF) (98,99). As mentioned previously, the expression of EGF and TGF- $\alpha$  is substantially down-regulated and the signal transduction pathways are impaired in microgravity (51–53,100–103). Interestingly, simulated microgravity enhances the activity of KGF (49). Therefore, any impairment of EGF expression that is caused by microgravity might be partially masked through a parallel upregulation of KGF.

PDGF is another factor that must be considered, as PDGF appears to be essential for normal wound healing (104,105). PDGF has two distinct roles in healing procedure: an early function to stimulate fibroblast proliferation and a later function to induce the myofibroblast phenotype (106). Akiyama *et al.* (54) found that the expression of PDGF in microgravity is 62% lower than the control ground samples. Furthermore, the early function of PDGF – the stimulation of fibroblast proliferation – is substantially diminished in this condition (55). Meanwhile, space flight-induced psychological stress may lead to downregulation of PDGF and its receptor in wound milieu (107).

Gap junctional intercellular communications have implicated to play an important role in wound healing (108). Following trauma, rapid immediate closure of gap junctions takes places, uncoupling damaged cells from uninjured ones (109). During reepithelialisation, gap junctions are temporarily lost on the surface of keratinocytes located in the leading edge of centripetal moving rim of epidermis (110). Also, fibroblasts derived from nodules that were excised from Dupuytren's contracture lesions show reduced levels of intracellular gap junctions compared with normal dermal fibroblasts (111). In contrast, promotion of intracellular gap junctions through daily injections of LiCl into polyvinyl alcohol sponge implanted into the wound milieu, enhanced the penetration of granulation tissue into the interstices of the sponge, increased the amount of connective tissue deposited in the surrounding capsule and promoted more organised collagen fibres (112). Liu et al. (113) reported that connexin 43 decreased significantly and distributed irregularly after simulated microgravity. On the contrary, Claassen and Spooner could not detect any alteration in channelling activity of cardiac gap junctions following short period microgravitational status (114).

Directional centripetal migration of keratinocytes into the wound bed is necessary for optimal wound healing. Keratinocytes express β2-adrenergic receptor. It has been shown that β-adrenergic receptor activation delays wound healing by preventing the organisation of the actin cytoskeleton and localisation of phosphoextracellular receptor kinase to the lamellipodial edge and its colocalisation with vinculin and thus leading to a considerable delay in reepithelialisation (115,116). Moreover, it has been suggested that β-adrenergic receptor antagonists promote wound reepithelialisation in chronic human skin wound (117). However, β-adrenergic receptor activation enhanced fibroblast proliferation and contraction and meanwhile decreased fibroblast-mediated collagen gel contraction, both of which are detrimental to wound healing (118,119). However,  $\beta$ 1- and  $\beta$ 2-adrenoceptor blockade impairs cutaneous wound healing, delineating the complicated role of sympathetic system in regulation of healing response (120). Simulation of microgravity through head-down bed rest induced increased responsiveness of sympathetic nervous system through β-adrenergic receptor sensitisation (121). Nonetheless, while sympathetic nervous activity was decreased during head-down bed rest, a similar finding was not observed during microgravity and thus head-down bed rest cannot be applied to simulate changes in sympathoadrenal activity during microgravity (122). Considering the existing controversy, investigation of microgravity-induced alteration of sympathetic activity and its effect on healing dermal wounds seems necessary.

Stem cells from various proximal and distal niches are involved in wound healing. It has been suggested that human mesenchymal stem cells (hMSCs) together with b-fibroblast growth factor accelerate cutaneous wound healing as the hMSCs transdifferentiate into the epithelium (123). The contribution of epidermal stem cells to repair of wounded epidermis is now evident (124,125). Moreover, adult bone-marrowderived mesenchymal stem cells home the sites of tissue injury and enhance the healing dynamics through differentiating into various cellular elements (126). It has been suggested that microgravity reduces proliferative as well as differentiation capabilities of human mesenchymal stem cells (127,128). However, newly emerging data contradict the proposed negative impacts of microgravity and underline its enhancement of proliferative activity of mesenchymal stem cells (129).

### CONCLUSION

It may be concluded that various stages of wound healing and sophisticated interactions between elements involved in healing response are modified in microgravitational status. However, future studies addressing the issue through practical approaches are necessary to understand the aforementioned alterations.

### REFERENCES

- 1 Stadelmann WK, Digenis AG, Tobin GR. Physiology and healing dynamics of chronic cutaneous wounds. Am J Surg 1998;176 Suppl 2A:26S–38S.
- 2 Diegelmann RF, Evans MC. Wound healing: an overview of acute, fibrotic and delayed healing. Front Biosci 2004;9:283–9.
- 3 Martin P. Wound healing-aiming for perfect skin regeneration. Science 1997;276:75–81.
- 4 Arquilla ER, Weringer EJ, Nakajo M. Wound healing: a model for the study of diabetic angiopathy. Diabetes 1976;25 (2 Suppl):811–9.
- 5 Kiecolt-Glaser JK, Marucha PT, Malarkey WB, Mercado AM, Glaser R. Slowing of wound healing by psychological stress. Lancet 1995;346:1194–6.
- 6 Marucha PT, Kiecolt-Glaser JK, Favagehi M. Mucosal wound healing is impaired by psychological stress. Psychosom Med 1998;60:362–5.
- 7 Bucalo B, Eaglstein WH, Falanga V. Inhibition of cell proliferation by chronic wound fluid. Wound Repair Regen 1993;1:181–6.
- 8 Pilcher BK, Wang M, Qin XJ, Parks WC, Senior RM, Welgus HG. Role of matrix metalloproteinases and their inhibition in cutaneous wound healing and allergic contact hypersensitivity. Ann N Y Acad Sci 1999;878:12–24.
- 9 Buckey JC. Space physiology. New York: Oxford University Press, 2006.
- 10 Nicogossian AE, Pool SL. Space physiology and medicine, 4th edn. Baltimore: Lippincott Williams & Wilkins, 2003.
- 11 Hinsbergh VWM, Collen A, Koolwijk P. Role of fibrin matrix in angiogenesis. Ann N Y Acad Sci 2001:936:421.
- 12 Laurens N, Koolwijk P, De Maat MPM. Fibrin structure and wound healing. J Thromb Haemost 2006;4:932.
- 13 Weibrich G, Kleis WKG, Hafner G, et al. Growth factor levels in platelet-rich plasma and correlation with donor age, sex and platelet count. J Craniomaxillofac Surg 2002;30:97.
- 14 Landesberg R, Roy M, Glickman RS. Quantification of growth factor levels using a simplified method of platelet-rich plasma gel preparation. J Oral Maxillofac Surg 2000;58:297.
- 15 Rowe WJ. The Apollo 15 space syndrome. Circulation 1998;97:119–20.
- 16 Cooke WH, Convertino VA. Cardiovascular consequences of weightlessness promote advances in clinical and trauma care. Curr Pharm Biotechnol 2005;6:285–97
- 17 Fuse A, Aoki Y, Sato T, Sunohara M, Takeoka H.

  Decreased platelet level in peripheral blood of

- mice by microgravity. Biol Sci Space 2002;16: 159–60.
- 18 Kalandarova MP. Changes in hematologic indicators in personnel testing during 370-day anti-orthostatic hypokinesia. Kosm Biol Aviakosm Med 1991;25:15–8.
- 19 Davis TA, Wiesmann W, Kidwell W, Cannon T, Kerns L, Serke C, Delaplaine T, Pranger A, Lee KP. Effect of spaceflight on human stem cell hematopoiesis and myelopoiesis. J Leukoc Biol 1996;60:69–76.
- 20 Dintenfass L. Experiment on "Discovery" STS 51-C aggregation of red cells and thrombocytes in heart disease, hyperlipidaemia and other conditions. Adv Space Res 1989;9:65-9.
- 21 Raffai G, Kocsis L, Meszaros M, Monos E, Dezsi L. Inverse-orthostasis may induce elevation of blood pressure due to sympathetic activation. J Cardiovasc Pharmacol 2006;47:287–94.
- 22 Watenpaugh DE. Fluid volume control during short-term space flight and implications for human performance. J Exp Biol 2001;204:3209–15.
- 23 McCuaig K, Lloyd CW, Gosbee J, Snyder WW. Simulation of blood flow in microgravity. Am J Surg 1992;164:119–23.
- 24 Stein TP. Space flight and oxidative stress. Nutrition 2002;18:867–71.
- 25 Markin A, Strogonova L, Balashov O, Polyakov V, Tigner T. The dynamics of blood biochemical parameters in cosmonauts during long-term space flights. Acta Astronaut 1998;42:247–53.
- 26 Kimzey SL, Ritzmann SE, Mengel CE, Fischer CL. Skylab experiment results: hematology studies. Acta Astronaut 1975;2:141–54.
- 27 Nunes CR, Roedersheimer MT, Simske SJ, Luttges MW. Effect of microgravity, temperature, and concentration on fibrin and collagen assembly. Microgravity Sci Technol 1995;8:125–30.
- 28 Roedersheimer MT, Bateman TA, Simske SJ. Effect of gravity and diffusion interface proximity on the morphology of collagen gels. J Biomed Mater Res 1997;37:276–81.
- 29 Carr ME Jr, Carr SL. Fibrin structure and concentration alter clot elastic modulus but do not alter platelet mediated force development. Blood Coagul Fibrinolysis 1995;6:79–86.
- 30 Braaten JV, Jerome WG, Hantgan RR. Uncoupling fibrin from integrin receptors hastens fibrinolysis at the platelet-fibrin interface. Blood 1994;83: 982–93.
- 31 Murciano JC, Harshaw D, Neschis DG, Koniaris L, Bdeir K, Medinilla S, Fisher AB, Golden MA, Cines DB, Nakada MT, Muzykantov VR. Platelets inhibit the lysis of pulmonary microemboli. Am J Physiol Lung Cell Mol Physiol 2002;282:529–39.
- 32 Collawn SS. Occlusion following laser resurfacing promotes reepithelialization and wound healing. Plast Reconstr Surg 2000;105:2180–9.
- 33 Campbell MR, Billica RD, Johnston SL 3rd. Surgical bleeding in microgravity. Surg Gynecol Obstet 1993;177:121–5.
- 34 Rafiq A, Hummel R, Lavrentyev V, Derry W, Williams D, Merrell RC. Microgravity effects on fine motor skills: tying surgical knots during

# **Key Points**

- it may be concluded that various stages of wound healing and sophisticated interactions between elements involved in healing response are modified in microgravitational status
- future studies addressing the issue through practical approaches are necessary to understand the aforementioned alterations

- parabolic flight. Aviat Space Environ Med 2006; 77:852–6.
- 35 Halloran BG, So BJ, Baxter BT. Platelet-derived growth factor is a cofactor in the induction of 1 alpha(I) procollagen expression by transforming growth factor-beta 1 in smooth muscle cells. J Vasc Surg 1996;23:767–73.
- 36 Janat MF, Liau G. Transforming growth factor beta 1 is a powerful modulator of platelet-derived growth factor action in vascular smooth muscle cells. J Cell Physiol 1992;150:232–42.
- 37 Lynch SE, Nixon JC, Colvin RB, Antoniades HN. Role of platelet-derived growth factor in wound healing: synergistic effects with other growth factors. Proc Natl Acad Sci USA 1987;84:7696–700.
- 38 Parekh T, Saxena B, Reibman J, Cronstein BN, Gold LI. Neutrophil chemotaxis in response to TGF-β isoforms (TGF-β1, TGF-β2, TGF-β3) is mediated by fibronectin. J Immunol 1994;152:2456–66.
- 39 Roberts AB, Sporn MB, Assoian RK, Smith JM, Roche NS, Wakefield LM, Heine UI, Liotta LA, Falanga V, Kehrl JH, Fauci AS. Transforming growth factor type beta: rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. Proc Natl Acad Sci U S A 1986;83:4167–71.
- 40 Czuwara-Ladykowska J, Gore EA, Shegogue DA, Smith EA, Trojanowska M. Differential regulation of transforming growth factor-beta receptors type I and II by platelet-derived growth factor in human dermal fibroblasts. Br J Dermatol 2001; 145:569–75.
- 41 Graves LM, Han J, Earp HS. Transactivation of the EGF receptor: is the PDGF receptor an unexpected accomplice? Mol Interv 2002;2:208–12.
- 42 Mccarthy DW, Downing MT, Brigstock DR, Luquette MH, Brown KD, Abad MS, Besner GE. Production of heparin-binding epidermal growth factor-like growth factor (HB-EGF) at sites of thermal injury in pediatric patients. J Invest Dermatol 1996;106:49–56.
- 43 Todd R, Donoff BR, Chiang T, Chou MY, Elovic A, Gallagher GT, Wong DT. The eosinophil as a cellular source of transforming growth factor alpha in healing cutaneous wounds. Am J Pathol 1991;138:1307–13.
- 44 Yu W, Naim JO, Lanzafame RJ. Expression of growth factors in early wound healing in rat skin. Lasers Surg Med 1994;15:281–9.
- 45 O'Kane S, Ferguson MWJ. Transforming growth factor βs and wound healing. Int J Biochem Cell Biol 1997;29:63–78.
- 46 Levine JH, Moses HL, Gold LI, Nanney LB. Spatial and temporal patterns of immunoreactive transforming growth factor β1, β2, and\_β3 during excisional wound repair. Am J Pathol 1993;143: 368–80.
- 47 Ansel JC, Tiesman JP, Olerud JE, Krueger JG, Krane JF, Tara DC, Shipley GD, Gilbertson D, Usui ML, Hart CE. Human keratinocytes are a major source of cutaneous platelet-derived growth factor. J Clin Invest 1993;92:671–8.
- 48 Antoniades HN, Galanopoulos T, Neville-Golden J, Kiritsy CP, Lynch SE. Injury induces in vivo

- expression of platelet-derived growth factor (PDGF) and PDGF receptor mRNAs in skin epithelial cells and PDGF mRNA in connective tissue fibroblasts. Proc Natl Acad Sci U S A 1991; 88:565–9.
- 49 Beer HD, Longaker MT, Werner S. Reduced expression of PDGF and PDGF receptors during impaired wound healing. J Invest Dermatol 1997; 109:132–8.
- 50 Whitby DJ, Ferguson MWJ. Immunohistochemical localization of growth factors in fetal wound healing. Dev Biol 1991;147:207–15.
- 51 Jessup JM, Frantz M, Sonmez-Alpan E, Locker J, Skena K, Waller H, Battle P, Nachman A, Weber ME, Thomas DA, Curbeam RL Jr, Baker TL, Goodwin TJ. Microgravity culture reduces apoptosis and increases the differentiation of a human colorectal carcinoma cell line. In Vitro Cell Dev Biol Anim 2000;36:367–73.
- 52 Rijken PJ, Boonstra J, Verkleij AJ, de Laat SW. Effects of gravity on the cellular response to epidermal growth factor. Adv Space Biol Med 1994;4:159–88.
- 53 Boonstra J. Growth factor-induced signal transduction in adherent mammalian cells is sensitive to gravity. FASEB J 1999;13:S35–42.
- 54 Akiyama H, Kanai S, Hirano M, Shimokawa H, Katano H, Mukai C, Nagaoka S, Morita S, Kumei Y, Akiyama H, Kanai S, Hirano M, Shimokawa H, Katano H, Mukai C, Nagaoka S, Morita S, Kumei Y. Expression of PDGF-β receptor, EGF receptor, and receptor adaptor protein Shc in rat osteoblasts during spaceflight. Mol Cell Biochem 1999;202:63–71.
- 55 Davidson JM, Aquino AM, Woodward SC, Wilfinger WW. Sustained microgravity reduces intrinsic wound healing and growth factor responses in the rat. FASEB J 1999;13:325–9.
- 56 Hart J. Inflammation. 1: its role in the healing of acute wounds. J Wound Care 2002;11:205.
- 57 Sylvia CJ. The role of neutrophil apoptosis in influencing tissue repair. J Wound Care 2003;12:13.
- 58 Riches DWH. Macrophage involvement in wound repair, remodeling, and fibrosis. In: Clark RAF, editor. The molecular and cellular biology of wound repair. New York: Plenum, 1996:95–142.
- 59 Taylor GR, Neale LS, Dardano JR. Immunologic analysis of U.S. space shuttle crew members. Aviat Space Environ Med 1986;57:213–7.
- 60 Ichiki AT, Gibson LA, Jago TL, Strickland KM, Johnson DL, Lange RD, Allebban Z. Effects of spaceflight on rat peripheral blood leukocytes and bone marrow progenitor cells. J Leukoc Biol 1996;60:37–43.
- 61 Meehan RT, Whitson P, Sams C. The role of psychoneuroendocrine factors on spaceflight-induced immunological alterations. J Leukoc Biol 1993:54:236–44.
- 62 Meehan RT, Neale LS, Kraus ET, Stuart CA, Smith ML, Cintron NM, Sams CF. Alteration in human mononuclear leukocytes following spaceflight. Immunology 1992;76:491–7.
- 63 Allebban Z, Ichiki AT, Gibson LA, Jones JB, Congdon CC, Lange RD. Effects of spaceflight on the

- number of rat peripheral blood leukocytes and lymphocyte subsets. J Leukoc Biol 1994;55:209–13.
- 64 Stowe RP, Sams CF, Mehta SK, Kaur I, Jones ML, Feeback DL, Pierson DL. Leukocyte subsets and neutrophil function after short-term spaceflight. J Leukoc Biol 1999;65:179–86.
- 65 Boxer LA, Allen JM, Baehner RL. Diminished polymorphonuclear leukocyte adherence. Function dependent on release of cyclic AMP by endothelial cells after stimulation of b-receptors by epinephrine. J Clin Invest 1980;66:268–74.
- 66 Miller ES, Koebel DA, Davis SA, Klein JB, McLeish KR, Goldwater D, Sonnenfeld G. Influence of suspension on the oxidative burst by rat neutrophils. J Appl Physiol 1994;76:387–90.
- 67 Meloni MA, Galleri G, Camboni MG, Pippia P, Cogoli A, Cogoli-Greuter M. Modeled microgravity affects motility and cytoskeletal structures. J Gravit Physiol 2004;11:197–8.
- 68 Noli C, Miolo A. The mast cell in wound healing. Vet Dermatol 2001;12:303–13.
- 69 Popchristova E, Mazgalova J. Some observations on mast cell numbers in steroid-treated keloids. Ann Burns Fire Disas 1996;9:168.
- 70 Theoharides TC, Spanos CP, Pang X, Alferes L, Ligris K, Letourneau R, Rozniecki JJ, Webster E, Chrousos G. Stress-induced intracranial mast cell degranulation. A corticotropin releasing hormone-mediated effect. Endocrinology 1995;136: 5745–50.
- 71 Theoharides TC, Singh LK, Boucher W, Pang X, Letourneau R, Webster E, Chrousos G. Corticotropin-releasing hormone induces skin mast cell degranulation and increased vascular permeability, a possible explanation for its proinflammatory effects. Endocrinology 1998;139:403–13.
- 72 Pippia P, Sciola L, Cogoli-Greuter M, Meloni MA, Spano A, Cogoli A. Activation signals of T lymphocytes in microgravity. J Biotechnol 1996; 47:215–22.
- 73 Walther I, Pippia P, Meloni MA, Turrini F, Mannu F, Cogoli A. Simulated microgravity inhibits the genetic expression of interleukin-2 and its receptor in mitogen-activated T lymphocytes. FEBS Lett 1998;436:115–8.
- 74 Morrow MA. Clinorotation differentially inhibits T-lymphocyte transcription factor activation. In Vitro Cell Dev Biol Anim 2006;42:153–8.
- 75 Simons DM, Gardner EM, Lelkes PI. Dynamic culture in a rotating-wall vessel bioreactor differentially inhibits murine T-lymphocyte activation by mitogenic stimuli upon return to static conditions in a time-dependent manner. J Appl Physiol 2006;100:1287–92.
- 76 Peana AT, Bennardini F, Buttu L, Pippia P, Meloni MA, Stuffler RG, Maccarrone M. Effect of simulated microgravity on PGE2-induced edema and hyperalgesia in rat paws: pharmacological data and biochemical correlates. J Gravit Physiol 2004;11:41–2.
- 77 Kumei Y, Shimokawa H, Katano H, Hara E, Akiyama H, Hirano M, Mukai C, Nagaoka S, Whitson PA, Sams CF. Microgravity induces prostaglandin E[2] and interleukin-6 production

- in normal rat osteoblasts: role in bone demineralization. J Biotechnol 1996;47:313–24.
- 78 Carter K. Growth factors: the wound healing therapy of the future? Br J Community Nurs 2003;8 Suppl 9:S15–23.
- 79 Licato LL, Grimm EA. Multiple interleukin-2 signaling pathways differentially regulated by microgravity. Immunopharmacology 1999;44: 273–9.
- 80 Berry WD, Murphy JD, Smith BA, Taylor GR, Sonnenfeld G. Effect of microgravity modeling on interferon and interleukin responses in the rat. J Interferon Res 1991;11:243–9.
- 81 Stein TP, Schluter MD. Excretion of IL-6 by astronauts during spaceflight. Am J Physiol 1994;266:E448–52.
- 82 Roberts AB, Sporn MB. Physiological actions and clinical applications of transforming growth factor-beta (TGF-beta). Growth Factors 1993;8:1.
- 83 Roberts AB, McCune BK, Sporn MB. TGF-beta: regulation of extracellular matrix. Kidney Int 1992;41:557.
- 84 Hall MC, Young DA, Waters JG, Rowan AD, Chantry A, Edwards DR, Clark IM. The comparative role of activator protein 1 and Smad factors in the regulation of Timp-1 and MMP-1 gene expression by transforming growth factor-beta 1. J Biol Chem 2003;278:10304.
- 85 Westerlind KC, Turner RT. The skeletal effects of spaceflight in growing rats: tissue-specific alterations in mRNA levels for TGF-beta. J Bone Miner Res 1995;10:843–8.
- 86 Zhang R, Supowit SC, Hou X, Simmons DJ. Transforming growth factor-β2 mRNA level in unloaded bone analyzed by quantitative in situ hybridization. Calcif Tissue Int 1999;64:522–26.
- 87 Carmeliet G, Nys G, Bouillon R. Microgravity reduces the differentiation of human osteoblastic MG-63 cells. J Bone Miner Res 1997;12:786–94.
- 88 Periyasamy S, Sanchez ER. Antagonism of glucocorticoid receptor transactivity and cell growth inhibition by transforming growth factor-beta through AP-1-mediated transcriptional repression. Int J Biochem Cell Biol 2002;34:1571–85.
- 89 Song CZ, Tian X, Gelehrter TD. Glucocorticoid receptor inhibits transforming growth factor-β signaling by directly targeting the transcriptional activation function of Smad3. Proc Natl Acad Sci U S A 1999;96:11776–81.
- 90 Sakai K, Mohtai M, Iwamoto Y. Fluid shear stress increases transforming growth factor beta 1 expression in human osteoblast-like cells: modulation by cation channel blockades. Calcif Tissue Int 1998;63:515–20.
- 91 Ashcroft GS, Yang X, Glick AB, Weinstein M, Letterio JL, Miezel DE, Anzano M, Greenwell-Wild T, Wahl SM, Deng C, Roberts AB. Mice lacking Smad3 show accelerated wound healing and an impaired local inflammatory response. Nat Cell Biol 1999;1:260–6.
- 92 Yarden Y. The EGFR family and its ligands in human cancer: signaling mechanisms and therapeutic opportunities. Eur J Cancer 2001;37 Suppl 4:3–8.

- 93 Stodcheck CM, Nanney LB, King LE Jr. Quantitative determination of EGF-R during epidermal wound healing. J Invest Dermatol 1992;9:645–9.
- 94 Wenczak BA, Lynch JB, Nenney LB. Epidermal growth factor receptor distribution in burn wounds. Implications for growth factor-mediated repair. J Clin Invest 1992;90:2392–401.
- 95 Greenhalgh DG. The role of growth factors in wound healing. J Trauma 1996;41:159–67.
- 96 Steed DL. Modifying the wound healing response with exogenous growth factors. Clin Plast Surg 1998;25:397–405.
- 97 Kim I, Mogford JE, Chao JD, Mustoe TA. Wound epithelialization deficits in the transforming growth factor-alpha knockout mouse. Wound Repair Regen 2001;9:386–90.
- 98 Luetteke NC, Qiu TH, Peiffer RL, Oliver P, Smithies O, Lee DC. TGF-deficiency results in hair follicle and eye abnormalities in targeted and waved-1 mice. Cell 1993;73:263–78.
- 99 Sato A, Hamazaki T, Oomura T, Osada H, Kakeya M, Watanabe M, Nakamura T, Nakamura Y, Koshikawa N, Yoshizaki I, Aizawa S, Yoda S, Ogiso A, Takaoki M, Kohno Y, Tanaka H. Effects of microgravity on c-fos gene expression in osteoblast-like MC3T3-E1 cells. Adv Space Res 1999;24:807–13.
- 100 Rijken PJ, de Groot RP, Kruijer W, de Laat SW, Verkleij AJ, Boonstra J. Identification of specific gravity sensitive signal transduction pathways in human A431 carcinoma cells. Adv Space Res 1992; 12:145–52.
- 101 Rijken PJ, de Groot RP, Kruijer W, Verkleij AJ, Boonstra J, de Laat SW. Altered gravity conditions affect early EGF-induced signal transduction in human epidermal A431 cells. ASGSB Bull 1992;5:77–82.
- 102 Rijken PJ, de Groot RP, van Belzen N, de Laat SW, Boonstra J, Verkleij AJ. Inhibition of EGF-induced signal transduction by microgravity is independent of EGF receptor redistribution in the plasma membrane of human A431 cells. Exp Cell Res 1993;204:373–7.
- 103 Martin A, Zhou A, Gordon RE, Henderson SC, Schwartz AE, Schwartz AE, Friedman EW, Davies TF. Thyroid organoid formation in simulated microgravity: influence of keratinocyte growth factor. Thyroid 2000;10:481–7.
- 104 Pierce GF, Tarpley JE, Tseng J, Bready J, Chang D, Kenney WC, Rudolph R, Robson MC, Vande Berg J, Reid P, Kaufman S, Farrell CL. Detection of platelet-derived growth factor (PDGF)-AA in actively healing human wounds treated with recombinant PDGF-BB and absence of PDGF in chronic nonhealing wounds. J Clin Invest 1995; 96:1336–50.
- 105 Clark RA. Regulation of fibroplasia in cutaneous wound repair. Am J Med Sci 1993;306:42–8.
- 106 Katz MH, Alvarez AF, Kirsner RS, Eaglstein WH, Falanga V. Human wound fluid from acute wounds stimulates fibroblast and endothelial cell growth. J Am Acad Dermatol 1991;25:1054–8.
- 107 Beer HD, Fässler R, Werner S. Glucocorticoidregulated gene expression during cutaneous wound repair. Vitam Horm 2000;59:217–39.

- 108 Goliger JA, Paul DL. Wounding alters epidermal connexin expression and gap junction-mediated intercellular communication. Mol Biol Cell 1995; 6:1491–501.
- 109 Bryan PJ, Franzea SE. Wound healing, cell communication and DNA synthesis during maginal disc regeneration in Drosophila. Dev Biol 1988;127: 197–208.
- 110 Lampe PD, Nguyen BP, Gil S, Usui M, Olerud J, Takada Y, Carter WG. Cellular interaction of integrin alpha3beta1 with laminin 5 promotes gap junctional communication. J Cell Biol 1998; 143:1735–47.
- 111 Moyer KE, Banducci DR, Graham WP, Ehrlich HP. Dupuytren's disease: changes in nodule and cord fibroblast physiology through in vitro aging. Plast Reconstr Surg 2002;110:187–93.
- 112 Moyer KE, Davis A, Saggars GC, Mackay DR, Ehrlich HP. Wound healing. The role of gap junctional communications in rat granulation tissue maturation. Exp Mol Pathol 2002;72:10–6.
- 113 Liu ZX, Ma TM, Yang HH, Wu DW, Wang DS, Zhang SJ. Impact of simulated microgravity on the expression and distribution of cardiac gap junction protein CX43. Space Med Med Eng (Beijing) 2003;16:448–51.
- 114 Claassen DE, Spooner BS. Effects of microgravity on liposome-reconstituted cardiac gap junction channeling activity. Biochem Biophys Res Commun 1989;161:358–62.
- 115 Pullar CE, Grahn JC, Liu W, Isseroff RR. ß2-adrenergic receptor activation delays wound healing. FASEB J 2006;20:76–86.
- 116 Chen J, Hoffman BB, Isseroff RR. Beta-adrenergic receptor activation inhibits keratinocyte migration via a cyclic adenosine monophosphate-independent mechanism. J Invest Dermatol 2002;119: 1261–8.
- 117 Pullar CE, Rizzo A, Isseroff RR. β-adrenergic receptor antagonists accelerate skin wound healing: evidence for a catecholamine synthesis network in the epidermis. J Biol Chem 2006;281: 21225–35.
- 118 Pullar CE, Isseroff RR. The β2-adrenergic receptor activates pro-migratory and pro-proliferative pathways in dermal fibroblasts via divergent mechanisms. J Cell Sci 2006;119:592–602.
- 119 Pullar CE, Isseroff RR. β2-adrenergic receptor activation delays dermal fibroblast-mediated contraction of collagen gels via a cAMP-dependent mechanism. Wound Repair Regen 2005;13:405–11.
- 120 Souza BR, Santos JS, Costa AMA. Blockade of  $\beta_1$ -and  $\beta_2$ -adrenoceptors delays wound contraction and re-epithelialization in rats. Clin Exp Pharmacol Physiol 2006;33:421.
- 121 Barbe P, Galitzky J, Thalamas C, Langin D, Lafontan M, Senard JM, Berlan M. Increase in epinephrine-induced responsiveness during microgravity simulated by head-down bed rest in humans. J Appl Physiol 1999;87:1614–20.
- 122 Christensen NJ, Heer M, Ivanova K, Norsk P. Sympathetic nervous activity decreases during head-down bed rest but not during microgravity. J Appl Physiol 2005;99:1552–7.

- 123 Nakagawa H, Akita S, Fukui M, Fujii T, Akino K. Human mesenchymal stem cells successfully improve skin-substitute wound healing. Br J Dermatol 2005:153:29–36.
- 124 Ito M, Liu Y, Yang Z, Nguyen J, Liang F, Morris RJ, Cotsarelis G. Stem cells in the hair follicle bulge contribute to wound repair but not to homeostasis of the epidermis. Nat Med 2005;11: 1351–4.
- 125 Morasso MI, Tomic-Canic M. Epidermal stem cells: the cradle of epidermal determination, differentiation and wound healing. Biol Cell 2005;97: 173–83.
- 126 Li H, Fu X, Ouyang Y, Cai C, Wang J, Sun T. Adult bone-marrow-derived mesenchymal stem cells

- contribute to wound healing of skin appendages. Cell Tissue Res 2006;326:725–36.
- 127 Merzlikina NV, Buravkova LB, Romanov YA. The primary effects of clinorotation on cultured human mesenchymal stem cells. J Gravit Physiol 2004;11: 193–4.
- 128 Zayzafoon M, Gathings WE, McDonald JM. Modeled microgravity inhibits osteogenic differentiation of human mesenchymal stem cells and increases adipogenesis. Endocrinology 2004;145:2421–32.
- 129 Yuge L, Kajiume T, Tahara H, Kawahara Y, Umeda C, Yoshimoto R, Wu SL, Yamaoka K, Asashima M, Kataoka K, Ide T. Microgravity potentiates stem cell proliferation while sustaining the capability of differentiation. Stem Cells Dev 2006;15:921–9.